

SYNTHESIS OF ENANTIOMERICALLY PURE (10R,11S)- (+)-JUVENILE HORMONES I AND II†

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Abstract -- Enantiomerically pure (+)-juvenile hormone I [methyl (2E,6E,10R,11S)-10,11-epoxy-3,11-dimethyl-7-ethyl-2,6-tridecadienoate; *Cecropia* JH = C₁₈ JH = JH I] and (+)-juvenile hormone II [methyl (2E,6E,10R,11S)-10,11-epoxy-3,7,11-trimethyl-2,6-tridecadienoate; C₁₇ JH = JH II] were synthesized employing the enantioselective reduction of 2-ethyl-2-methyl-1,3-cyclohexanedione by a yeast (*Pichia terricola* KI 0117) as the key-step.

Since Rölller's discovery of juvenile hormone I (JH I) in 1967,¹ four additional juvenile hormones were isolated and identified as shown in Fig. 1 (JH II,² JH III,³ JH 0,⁴ and 4-Me JH I⁵). Importance of the juvenile hormones in both basic and applied entomological research was well appreciated from the beginning, and an enormous amount of works were published in the past by chemists and entomologists.⁶ Due to the limited amount of the juvenile hormones available from insects, their absolute configuration was difficult to be determined. Indeed still at present the absolute configuration of both JH 0 and 4-Me JH I remains obscure. In 1970 Meyer and Hanzmann were able to isolate 1.372 mg of a mixture of JH I and JH II (90.2:9.8 mol%) and showed it to be dextrorotatory, $[\alpha]_D \approx +7^\circ$ (CHCl₃).⁷ They then proposed 10R,11S configuration to JH I and JH II by the application of Horeau's method of determination of absolute configuration.⁸ Subsequently two independent syn-

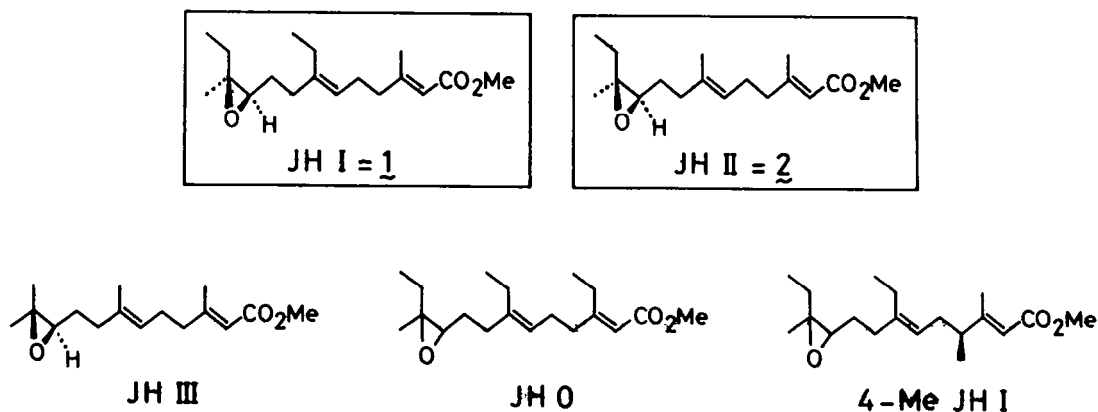


Fig. 1. Structures of juvenile hormones.

† Synthesis of compounds with juvenile hormone activity -- 26. Part 25, K. Mori and H. Mori, *Tetrahedron*, in the press. The experimental part of this work was taken from the forthcoming M. Sc. thesis of M. F. (March, 1988).

theses of the optically active form of JH I were achieved by Loew and Johnson⁹ and by Faulkner and Petersen.¹⁰ By using Johnson's sample of (+)-JH I, Nakanishi *et al.* proposed its $10R,11S$ configuration basing on physicochemical measurements.¹¹ The definite proof of the proposed $10R,11S$ configuration was provided by Faulkner's synthesis, because it started from (S)-(+)-2,2-dimethoxy-3-methyl-3-pentanol to furnish $(10R,11S)$ -(+)-JH I.¹⁰ Although these syntheses were decisive in settling the stereochemical argument on (+)-JH I, they did not give enantiomerically pure (+)-JH I. In Johnson's case their enantiomers of JH I were of 80-90% e.e.,⁹ while in the case of Faulkner the products were neither diastereomerically nor enantiomerically pure.¹⁰ In 1985 Prestwich and Wawrzęńczyk achieved the third synthesis of (+)-JH I by employing the Sharpless asymmetric epoxidation.¹² Their product was of high enantiomeric purity (~95% e.e.), but the Sharpless epoxidation, in our experience, never gave a product of 100% e.e. In the case of (+)-JH II, there exists only one synthesis of it, utilizing a microbial kinetic resolution process.¹³ The enantiomeric purity of the product, however, was not determined.¹³ Herein we describe the synthesis of both (+)-JH I and (+)-JH II in enantiomerically pure state.

Preparation of the chiral building block (2S,3S)-5a.

Fig. 2 shows our synthetic plan, which is an extension of the strategy employed in our recent synthesis of (R)-(+)-JH III.¹⁴ If it is possible to prepare pure hydroxy ketone A, then Baeyer-Villiger oxidation of A after protecting the OH group will give lactone B. Conversion of B to acetonide ester C will be followed by its modification to alkyne D. The alkyne D will give diene esters E (R=Et and R=Me) by the sequence developed for the synthesis of (+)-JH III.¹⁴ Finally the diene esters E will give (+)-JH I and (+)-JH II, respectively, by the standard procedure.¹⁴

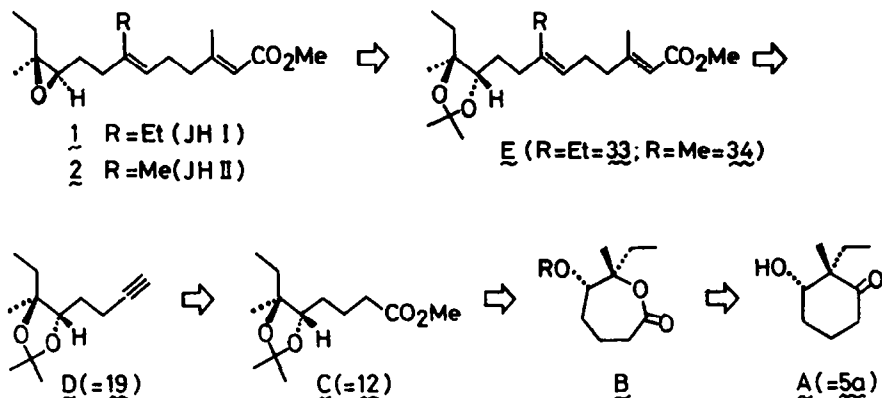


Fig. 2. Synthetic plan for (+)-JH I and (+)-JH II.

The first problem to be solved was therefore to find an efficient preparative method for enantiomerically pure A. In Fig. 3 is summarized our preparation of the chiral building block A [= (2S,3S)-5a]. Ethylation of 2-methylcyclohexane-1,3-dione (3) with EtI in the presence of Triton B in MeOH gave a mixture of the desired 4a and ethyl enol ether of 3. After hydrolyzing the enol ether with dil HCl, the product 4a (15% yield) was separated from the recovered 3 (76% recovery). By repeating this process, 4a was prepared in 63% yield from 3.^{cf.15} Our task was then to find a method to realize an efficient conversion of 4a to 5a (=A). Reduction of 4b with baker's yeast was known to give (S)-3-hydroxy-2,2-dimethylcyclohexanone (98-99% e.e.) in good yield.¹⁶ In the case of 4c, however, its reduction with baker's yeast gave a diastereomeric mixture of 7A and 7B.¹⁷ It was therefore difficult to predict the steric course of the biochemical reduction of 4a beforehand, and we began the screening experiments to find an appropriate strain of microorganism. The final result is summarized in Table 1, which deserves further explana-

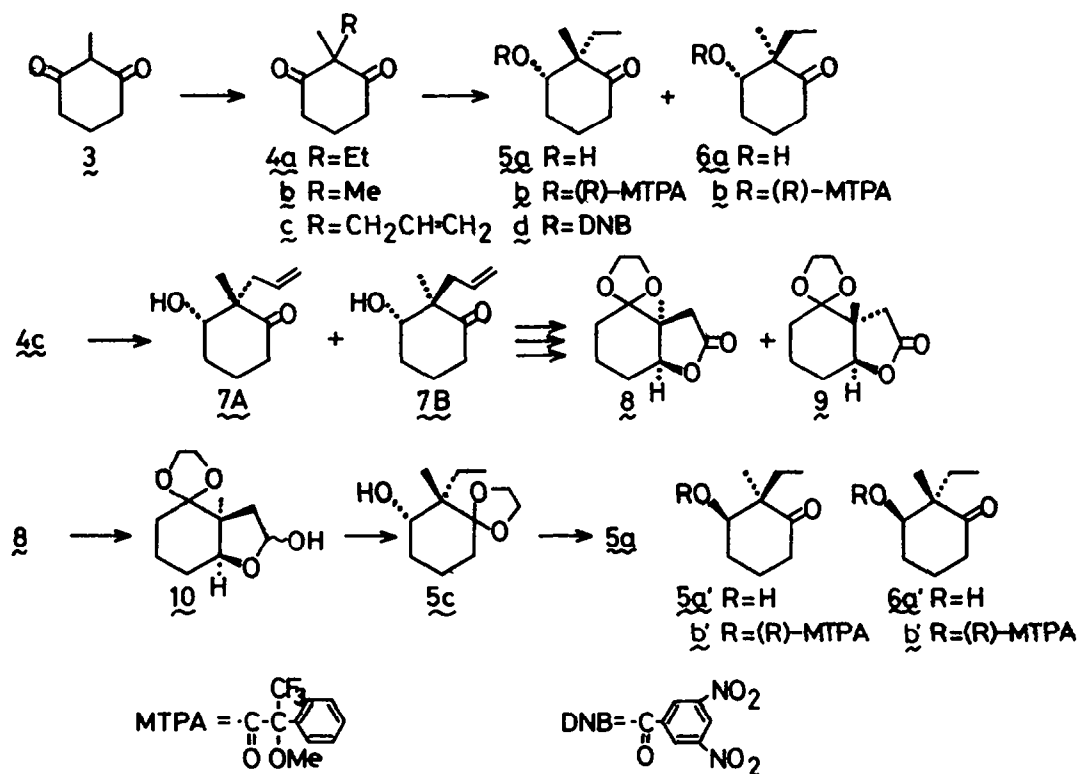


Fig. 3. Preparation of the chiral building block (2*S*,3*S*)-**5a**.

Table 1. Reduction of the prochiral ketone **4a** with yeast to give (2*S*,3*S*)-**5a**.

Yeast strain	Ratio of the generated hydroxy ketones 5a : 6a	Enantiomeric purity at C-3 (% e.e.)
<i>Saccharomyces cerevisiae</i> (baker's yeast)	1 : 2.7	99
<i>Saccharomyces bailii</i> KI 0116	1 : 1.5	73
<i>Pichia terricola</i> KI 0117	>99 : <1	99

tion as detailed below.

Our first choice was to employ baker's yeast, *Saccharomyces cerevisiae*. The reduction yielded a mixture of two ketols in a ratio of 1:2.7 as judged by the ^1H NMR analysis where a pair of Me singlets was observed. In analogy with Brooks's result,¹⁷ these two ketols were assumed to be **5a** and **6a**. Because **5a** was inseparable from **6a** by conventional TLC, the mixture was converted to the corresponding mixture of ethylene acetals **5c** and **6c** by treatment with ethylene glycol and *p*-TsOH in benzene under reflux. The acetals **5c** and **6c** were separable by SiO_2 chromatography. The two acetals **5c** and **6c** gave back the parent

ketones when treated with *p*-TsOH in acetone. The ^1H NMR spectra of the two ketones were readily distinguishable. It was difficult, however, to tell which was which (5a or 6a). Unambiguous derivation of 5a from a compound of known absolute configuration must therefore be the only way to assign absolute configuration to the reduction products generated by baker's yeast. The hydroxy ketone 7A was used for this purpose, because of its known absolute configuration.¹⁷ A mixture of 7A and 7B was submitted to a multi-step conversion to give a mixture of lactones 8 and 9.¹⁸ These two lactones were separated by chromatography, and their stereochemistries were unambiguously assigned on the basis of the magnitude of the J -value of the signal due to the angular -CHOCO- proton.^{18†} Reduction of 8 with $(i\text{-Bu})_2\text{AlH}$ gave lactol 10, which was reduced with N_2H_4 and KOH under the Huang Minlon condition to give 5c. Treatment of 5c with *p*-TsOH in acetone yielded 5a, whose ^1H NMR spectrum as well as the sign of its specific rotation was identical to that of the ketone derived from the less polar ethylene acetal. That ketone 5a turned out to be the minor product of the yeast reduction. Thus the less polar ethylene acetal was 5c, while the more polar one was 6c. It was now clear that the reduction with baker's yeast gave the desired 5a only as the minor product.

Although both of the above mentioned samples of 5a were dextrorotatory, the magnitude of their $[\alpha]_D$ values was different. Namely, the hydroxy ketone 5a derived from 8 exhibited $[\alpha]_D^{23} +77.1^\circ$ (CHCl_3), while the $[\alpha]_D$ value of 5a derived from 5c of yeast-reduction origin was only $+32.7^\circ$. The both samples of 5a were converted to the corresponding α -methoxy- α -trifluoromethylphenylacetates (MTPA esters),¹⁹ and analyzed by HPLC. The MTPA ester 5b of 5a originating from 8 was shown to be of 98.3% diastereomeric purity. However, the MTPA ester 5b of 5a derived from 5c (of yeast-reduction origin) was a mixture of two diastereomers (5b and 5b') in a ratio of 7:3, indicating the enantiomeric purity of this sample of 5a to be 40% e.e. In the course of acetalization of 5a to 5c employing *p*-TsOH and ethylene glycol in refluxing benzene, partial racemization apparently took place at C-3 due to a retroaldol-aldol process. After directly acylating the hydroxy ketone mixture (5a and 6a) produced by the reduction of 4a with baker's yeast, the resulting mixture of MTPA esters 5b, 5b', 6b and 6b' was analyzed by HPLC. Both 5a and 6a were found to be of 99% e.e. in accord with the similar study published by Brooks *et al.*¹⁷ The rather tedious and troublesome work as described above taught us the method how to estimate the enantio- and diastereoselectivity of the biochemical reduction of 4a.

As the result with baker's yeast was not so promising, we tried other yeasts. *Saccharomyces bailii* KI 0116 was known to reduce ethyl acetoacetate to ethyl (*S*)-3-hydroxybutanoate of 96% e.e.²⁰ When this yeast was employed for the reduction of 4a, neither the diastereoselectivity nor the enantioselectivity could be improved. Thus the ratio 5a to 6a in this reduction was 1:1.5 with the enantiomeric purity of 73% e.e. at C-3.

The next choice was *Pichia terricola* KI 0117.²¹ Fortunately, this yeast achieved the reduction with satisfactory selectivity. Reduction of the diketone 4a with *P. terricola* KI 0117 gave in 86% yield only the desired hydroxy ketone 5a of 99.5% e.e. To enhance the enantiomeric purity of 5a to 100% e.e., it was converted to the corresponding 3,5-dinitrobenzoate 5d. Highly crystalline nature of 5d allowed us to purify it by recrystallization giving pure 5d. Regeneration of 5a from 5d by hydrolysis was apt to accompany partial racemization at C-3 by the retroaldol-aldol mechanism. It was therefore impossible to check directly the enantiomeric purity of 5d at this stage. The enantiomeric purity of our intermediate was checked in a later stage on 14 in Fig. 4. The most important problem of selective reduction of 4a to 5a was thus solved by the help of the yeast, *P. terricola* KI 0117.

† In the case of 8, a 1H signal was observed at $\delta = 4.35$ with $W_{H/2} = 8$ Hz (eq H), while in the case of 9, a 1H signal was observable at $\delta = 4.03$ with $W_{H/2} = 18$ Hz (ax H).

Synthesis of the common intermediate 21 leading to both JH I and II.

The next phase of this work was to convert pure **5d** to the common intermediate **21**, which would lead to both JH I and JH II. Baeyer-Villiger oxidation of **5d** with MCPBA in CH_2Cl_2 gave crystalline lactone **11** quantitatively. Treatment of **11** with K_2CO_3 in MeOH to effect methanolysis of **11** to the corresponding dihydroxy ester was followed by acetone formation with $\text{Me}_2\text{C}(\text{OMe})_2$ and *p*-TsOH to furnish acetone ester **12** in 90% yield from **5d**. To check the diastereomeric purity of **12**, it was analyzed by capillary GLC to show a single peak. In its ^1H NMR spectrum, the 3H singlet due to C-6 Me appeared at δ 1.06.^{cf.22} To be sure about the satisfactory diastereomeric purity of **12**, a reference sample was prepared as follows, which contained both (\pm)-**12** and its diastereomer (\pm)-**13**. The crude **5a** obtained by the reduction with *P. terricola* KI 0117 was treated with aq KOH-MeOH to effect retroaldol-aldol reaction. The resulting mixture of (\pm)-**5a** and (\pm)-**6a** was acetylated and the mixture of acetoxy ketones was converted to a mixture of (\pm)-**12** and (\pm)-**13** as described above for the conversion of **5d** to **12**. These two diastereoisomers [(\pm)-**12** and (\pm)-**13**] were inseparable by capillary GLC. Thus the GLC analysis of **12** could not be the proof of its homogeneity. However, in the ^1H NMR spectrum of the mixture of (\pm)-**12** and (\pm)-**13**, two signals due to C-6 Me were observable, one at δ 1.06 and the other at δ 1.18.^{cf.22} The optically active **12** was therefore thought to be diastereomerically pure within the limit of the experimental error of the ^1H NMR measurement.

The next step was the reduction of **12** with LAH to furnish **14** in 90% yield. The enantiomeric purity of **14** was rigorously estimated as follows. Deprotection of the acetone group of **14** with hot aq AcOH yielded the parent triol, which was acylated with (*R*)- or (*S*)-MTPA-Cl. The resulting bis-(*R*)- and bis-(*S*)-MTPA esters **15** were clearly distinguishable by HPLC analysis. Each of them showed a single peak upon HPLC, confirming the ~100% enantiomeric purity of **14**.

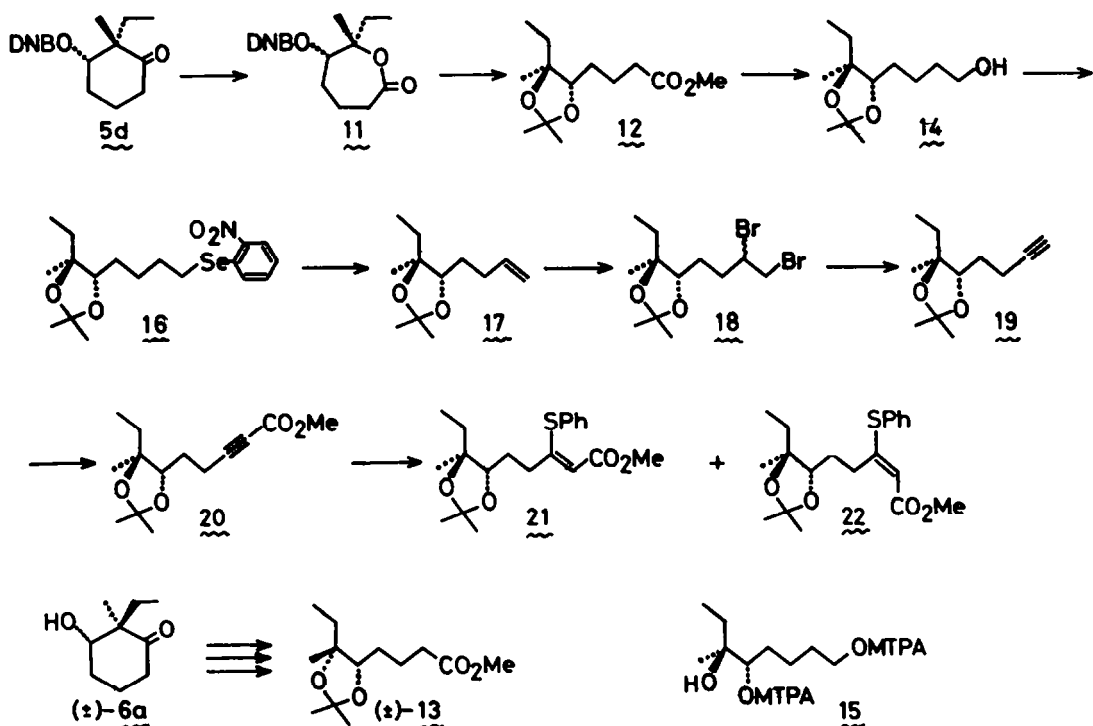


Fig. 4. Synthesis of the common intermediate **21**.

The remaining steps from 14 to 21 were almost same as those employed in our synthesis of (+)-JH III.¹⁴ Treatment of 14 with $\text{O}-(\text{O}_2\text{N})\text{C}_6\text{H}_4\text{SeCN}$ and $(n\text{-Bu})_3\text{P}$ in THF²³ gave selenide 16, which was immediately oxidized with H_2O_2 to give olefin 17 in 79% yield from 14. Bromination of 17 with Br_2 in CCl_4 furnished dibromide 18, which was dehydrobrominated with NaNH_2 in liq NH_3 to alkyne 19 in 50% yield from 17. According to Mukaiyama *et al.*,²⁴ 19 was methoxycarbonylated with $n\text{-BuLi}$ and ClCO_2Me in THF to give 20 in 86% yield. Treatment of 20 with PhSNa in MeOH afforded, after chromatographic separation, the desired common intermediate 21 in 76% yield together with the unwanted 22 (11% yield).

Synthesis of juvenile hormones I and II.

The third stage of this work was the preparation of (+)-JH I and (+)-JH II from the common intermediate 21. Substitution of the PhS group of 21 with Et would lead to JH I 1, and that with Me would yield JH II 2. Accordingly, treatment of 21 with EtMgBr or MeMgBr in the presence of CuI in THF at -65°C ²⁴ gave 23 (80% yield) or 24 (81% yield). These esters 23 and 24 were separately reduced with DIBALH to give alcohols 25 (quantitative) and 26 (90% yield), respectively. These were converted quantitatively to the corresponding bromides 27 and 28 by the method of Stork *et al.*²⁵ Alkylation of the dianion²⁶ of $\text{MeCOCH}_2\text{CO}_2\text{Me}$ with 27 and 28 gave 29 and 30, respectively. Their corresponding enol phosphates 31 and 32 were treated with Me_2CuLi to give 33 and 34.^{14, 26}

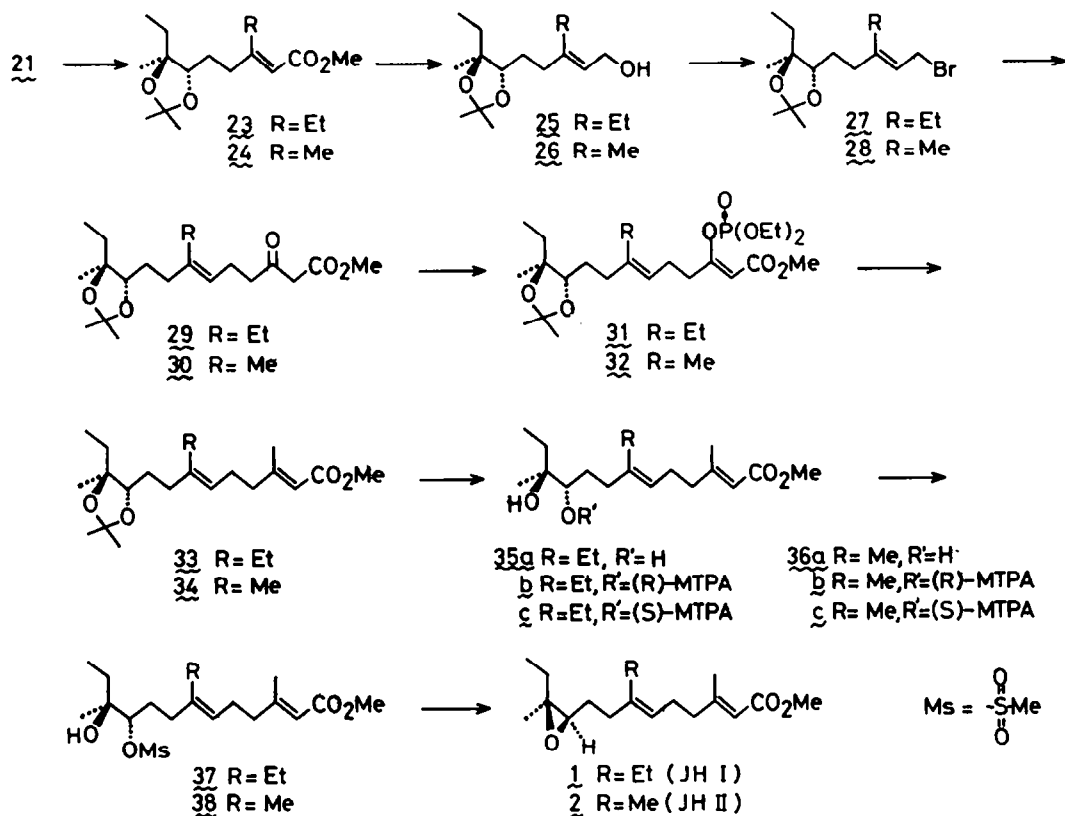


Fig. 5. Synthesis of JH I and JH II.

The final task of the epoxy-ring formation was executed as reported by us in the synthesis of (+)-JH III¹⁴ so as to avoid partial racemization at C-10. Hydrolytic removal of the acetonide protective group of 33 and 34 by treatment with hot aq AcOH gave 35a (99% yield) and 36a (quantitative), respectively. The enantiomeric purity of both 35a and 36a

was estimated to be ~100% by the HPLC analysis of the corresponding MTPA esters, 35b, 35c, 36b and 36c. Mesylation of 35a and 36a with methanesulfonic anhydride and Et₃N¹⁴ gave 37 and 38, respectively. These were treated with NaOMe in MeOH to give (+)-JH I [1, 78% from 35a; [α]_D²³ +14.9° (CHCl₃); [α]_D²³ +14.5° (MeOH); lit.⁹ [α]_D²⁰ +12.2° (CHCl₃)] and (+)-JH II [2, 66% yield from 36a; [α]_D^{24.5} +17.6° (MeOH); lit.¹³ [α]_D +11.7° (MeOH)], respectively.

As we rigorously checked the 100% enantiomeric purity of the dihydroxy esters 35a and 36a, the enantiomeric purity of our 1 and 2 must be 100%. Any racemization at C-10 would generate the diastereomers of JH I and JH II. To prove the high diastereomeric purity of our (+)-JH I and (+)-JH II, their ¹H NMR (400 MHz) and ¹³C NMR (25 MHz) spectra were carefully examined. Fig. 6 shows the NMR spectra of our JH I and JH II. These are the first published 400 MHz ¹H NMR and ¹³C NMR spectra of 1 and 2. The spectra clearly supported their 100% diastereomeric purity.^{27,28} Comparison of the 400 MHz ¹H NMR spectra of JH I and JH II as shown in Fig. 6 with the published ¹H NMR chart of JH I²⁹ and that of JH II^{2a} is quite instructive to show the remarkable improvement in NMR techniques in these twenty years.

In summary, the pure naturally occurring enantiomers of JH I and JH II were synthesized in 19 steps from the chiral building block 5a in 2.7% and 1.2% overall yield, respectively. Biochemical reduction of a prochiral diketone was again proved to be a useful method in preparing a chiral building block.

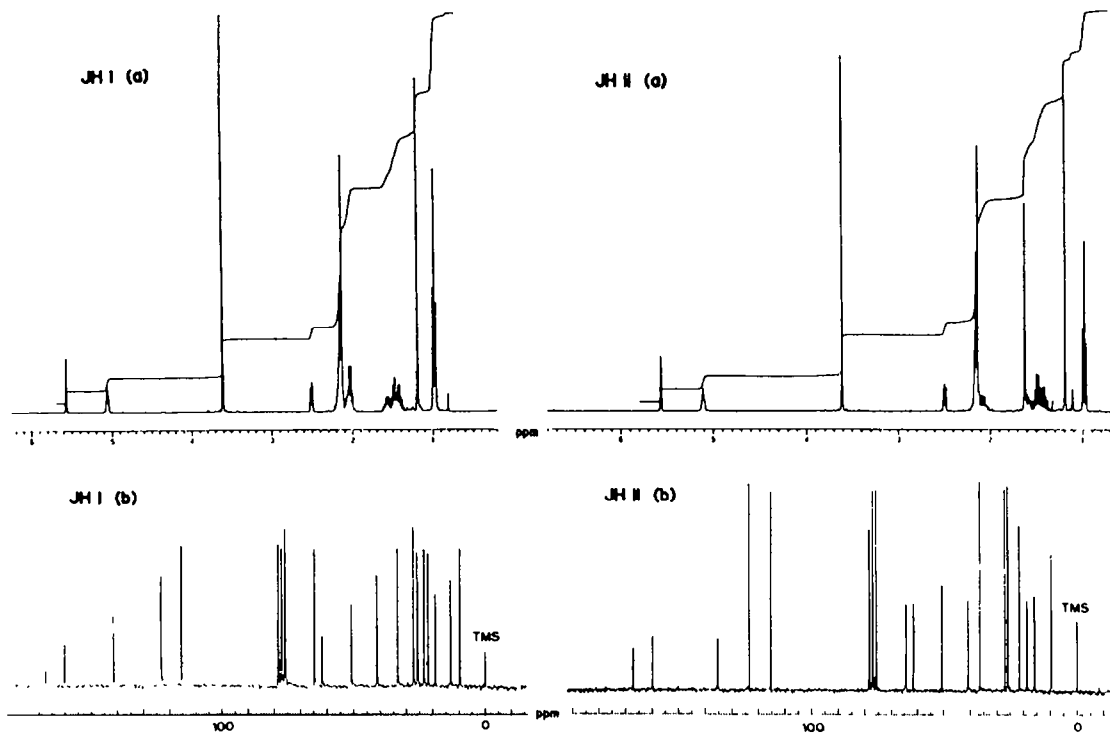


Fig. 6. NMR spectra of (+)-JH I and (+)-JH II.

(a) ¹H NMR Spectra (400 MHz, CCl₄).

(b) ¹³C NMR Spectra (25 MHz, CDCl₃).

EXPERIMENTAL

All bp's and mp's were uncorrected. IR spectra were measured as films for oils or as KBr discs for solids on a Jasco IRA-102 spectrometer. ¹H NMR spectra were recorded with TMS as an internal standard at 60 MHz on a Hitachi R-24A spectrometer or at 100 MHz on a JEOL JNM FX-100 spectrometer or at 400 MHz on a JEOL JNM GX-400 spectrometer. ¹³C NMR spectra were measured with TMS as an internal standard as CDCl₃ soln at 25 MHz on a Jeol JNM FX-100 spectrometer. Optical rotations were measured on a Jasco DIP 140 polarimeter. Merck Kieselgel 60 Art 7734 (particle size 0.063-0.200 mm) was

used for column chromatography. HPLC analyses were performed on Nucleosil® 50-5 (25 cm x 4.6 mm) as a column by the detection at 254 nm. The water used for the cultivation of yeast was deionized.

2-Ethyl-2-methylcyclohexane-1,3-dione 4a.^{cf.15} To a soln of 3 (37.8 g, 300 mmol) in MeOH (390 ml) was added a soln of Triton B (126 ml, 40% soln in MeOH) followed by EtI (51 g, 327 mmol) dropwise at room temp. The mixture was stirred and heated under reflux for 5 h. It was then cooled to room temp and MeOH was almost removed by evaporation. The residue was poured into N HCl soln and stirred for 10 min at room temp. Then the mixture was filtered and the residual solid on the filter was washed with EtOAc and dried under reduced pressure to give 28.6 g (75.7% recovery) of 3. The combined filtrate and washings were diluted with EtOAc and washed with 5% Na₂S₂O₃ soln, 20% K₂CO₃ soln and brine, dried (MgSO₄) and concentrated *in vacuo*. The residual oil was chromatographed over SiO₂ (100 g). Elution with *n*-hexane-EtOAc (20:1) gave 7.1 g (0.046 mmol, 15% yield, 63% yield based on the consumed 3) of 4a, ν_{\max} 3440 (w), 1730 and 1700 (s) cm⁻¹, δ (CCl₄) 0.73 (3H, t, J=7 Hz), 1.10 (3H, s), 1.40-2.20 (4H, m), 2.38-2.75 (4H, m).

Cultivation of *Saccharomyces bailii* KI 0116 or *Pichia terricola* KI 0117.²⁰ Two loops of *S. bailii* KI 0116 or *P. terricola* KI 0117 were inoculated into a 100 ml of medium containing malt extract (20 g), peptone (1 g), glucose (20 g) in water (1000 ml) placed in a 500 ml-shaking flask. This was cultivated at 37°C for 36 h on a reciprocal shaker (100 cpm). Two batches of the seed culture were added into a 1800 ml of medium with the same ingredient in a 5000 ml-Erlenmeyer cultivating flask with two internal projections for aeration. After cultivating at 37°C for 36 h on a gyratory shaker (100 rpm), the wet cells of yeast (ca. 10 g of *S. bailii* KI 0116 or ca. 22 g of *P. terricola* KI 0117) were harvested by centrifugation (3000 rpm) for 5 min.

Reduction of 4a with baker's yeast. An emulsion of 2-ethyl-2-methylcyclohexane-1,3-dione 4a (0.5 g) in Triton X-100 (0.2%, 10 ml) was treated with the dry cells of baker's yeast (Oriental Yeast Industries, 7 g) in sucrose soln (total 25g/100 ml water) at 30°C for 24 h. A usual workup and purification gave 0.26 g (1.7 mmol, 51%) of ketol; δ (100 MHz, CDCl₃) 1.12 and 1.15 (total 3H, each s, ca. 2:1). This was shown to be a ca. 1:2.7 mixture of two diastereomers.

Preparation of authentic samples of 5a and 6a. A mixture of 5a and 6a [obtained by the reduction with baker's yeast as above (0.70 g, 4.5 mmol)], ethylene glycol (1.5 g, 24 mmol) and *p*-TsOH·H₂O (8 mg) in benzene (15 ml) was stirred and heated under reflux for 15 h. After cooling, the mixture was poured into sat NaHCO₃ soln and extracted with ether. The ether soln was washed with brine, dried (MgSO₄) and concentrated *in vacuo* to give 0.93 g (quantitative) of a diastereomeric mixture of 5c and 6c. This was chromatographed over SiO₂ (70 g). Elution with *n*-hexane-EtOAc (20:1) gave 0.30 g of the less polar isomer, TLC [SiO₂, Merck Art 5715; developed with *n*-hexane-EtOAc (3:1)] Rf 0.42; ν_{\max} (film) 3550 (s), (CCl₄ soln) 3580 (m) cm⁻¹; the IR data suggested the presence of the intramolecular hydrogen bonding. Further elution gave a diastereomeric mixture (0.15 g). Further elution gave 0.40 g of the more polar isomer; TLC (same condition) Rf 0.33; ν_{\max} (film) 3475 (br,m), (CCl₄ soln) 3580 (s) cm⁻¹; this IR data showed the lack of intramolecular hydrogen bonding. In the case of 6c, internal hydrogen bonding causes an unfavorable 1,2-diaxial orientation of ethyl and hydroxyl group. Therefore the less polar isomer was tentatively assigned to be 5c, while the more polar isomer was to be 6c.

5c and 6c were separately deprotected to give 5a and 6a, respectively. 5a, $[\alpha]_D^{25} +32.7^\circ$ (c=1.00, CHCl₃); δ (100 MHz, CDCl₃) 0.89 (3H, t, J=8 Hz), 1.15 (3H, s), 3.72 (1H, dd, J=6, 9 Hz). 6a, $[\alpha]_D^{23.5} -25.0^\circ$ (c=1.00, CHCl₃); δ (100 MHz, CDCl₃) 0.87 (3H, t, J=8 Hz), 1.12 (3H, s), 3.92 (1H, m). The product obtained by the reduction with baker's yeast was therefore supposed to be a 1:2.7 mixture of 5a and 6a.

Determination of the optical purity. A mixture of 5a, 5a', 6a and 6a' was prepared as follows; to a soln of 5a (25 mg, 0.16 mmol) in MeOH (5 ml) was added 2 N KOH (0.1 ml) and the mixture was stirred for 2 h at room temp. Then the mixture was poured into water and extracted with ether. The ether soln was washed with brine, dried (MgSO₄) and concentrated *in vacuo* to give 20 mg (80%) of a mixture of 5a, 5a', 6a and 6a'. This mixture was converted to the corresponding (R)-MTPA esters. HPLC analysis [eluent: *n*-hexane-THF (30:1), 1 ml/min] showed three peaks; Rt 24.0 min (47.6%), 26.8 min (28.6%), 28.0 min (23.8%).

Authentic 5a and 6a were converted to the corresponding MTPA esters, respectively. HPLC analysis of MTPA ester of 5a: Rt 24.0 min (30.0%), 26.8 min (70.0%). MTPA ester of 6a: Rt 24.0 min (25.6%), 28.0 min (74.4%). Therefore the optical purity of authentic 5a and 6a was determined to be 40.0% e.e. and 48.8% e.e., respectively.

In the same manner, reduction product with baker's yeast was converted to the corresponding MTPA esters. HPLC analysis: Rt 24.0 min (0.5%), 26.8 min (26.6%), 28.0 min (72.9%). The optical purity was determined to be 99.0% e.e. The discrepancy of the optical purities as above was supposed to be due to the partial racemization or epimerization during acetalization.

Reduction of 4a with *S. bailii* KI 0116 and *P. terricola* KI 0117.

S. bailii KI 0116: Diketone 4a (0.5 g) was reduced with the wet cells of *S. bailii* KI 0116 (10 g) in glucose soln (total 15 g/100 ml water) at 37°C for 24 h to give 0.2 g of ketol; δ (100 MHz, CDCl₃) 1.12 and 1.15 (total 3H, each s, ca. 1.5:1). This was shown to be a ca. 1:1.5 mixture of 5a and 6a.

HPLC: (same condition); Rt 24.0 min (13.5%), 26.8 min (35.0%), 28.0 min (51.5%).

Pichia terricola KI 0117: Diketone 4a (0.5 g) was reduced with the wet cells of *P. terricola* KI 0117 (22 g) in glucose soln (total 15 g/100 ml water) at 37°C for 24 h to give 0.3 g of ketol; δ (100 MHz, CDCl₃) 1.15 (3H, s). Therefore reduction with *P. terricola* gave only one diastereomer.

HPLC: (same condition); Rt 24.0 min (0.3%), 26.8 min (99.5%), 28.0 min (0.2%). Therefore the ketol 5a obtained by *P. terricola* KI 0117 reduction was almost both enantiomerically (99.4% e.e.) and diastereomerically (99.6% d.e.) pure.

Determination of the absolute configuration of 5a.

1) (2R,3S,7S)-4,4-Ethylenedioxy-2-hydroxy-3b-methyloctahydrobenzofuran 10. According to Brooks *et al.*¹⁷ (2R,3S)-2-allyl-3-hydroxy-2-methylcyclohexanone (a mixture of 7A and 7B) was prepared by baker's yeast reduction of 2-allyl-2-methylcyclohexane-1,3-dione 4c. From this ketol, lactones (3bS,7bS)-4,4-ethylenedioxy-3b-methyl-2-oxo-octahydrobenzofuran 8 and (3bR,7bS)-isomer 9 were obtained *via* several steps. 8: m.p. 90-91°C; $[\alpha]_D^{21} -27.9^\circ$ (c=1.97, CHCl₃); ν_{\max} 1785 (s) cm⁻¹; δ (60 MHz, CDCl₃) 1.25 (3H, s), 4.35 (1H, deformed t, $\nu_{H/2}$ =8 Hz); TLC [SiO₂, developed with *n*-hexane-EtOAc (1:1)] Rf 0.3. 9 (oil): $[\alpha]_D^{21} -27.5^\circ$ (c=2.03, CHCl₃); ν_{\max} 1790 (s) cm⁻¹; δ (60 MHz, CDCl₃) 1.05 (3H, s), 4.03 (1H, dd, $\nu_{H/2}$ =18 Hz); TLC (same condition) Rf 0.4. Comparison of NMR spectra revealed that 8 had *cis*-configuration as well as 9 had *trans*-configuration, respectively. To a soln of 8 (540 mg, 2.55 mmol) in dry toluene (20 ml) was added a soln of DIBAL-H (3.5 ml, 1 mol/l

in toluene, 3.5 mmol) at -50°C under Ar and the mixture was stirred for 30 min at -60°C. Then the reaction was quenched by adding sat potassium sodium tartrate tetrahydrate soln (5 ml) and the mixture was stirred for 30 min at room temp. Then the mixture was poured into brine and extracted with ether. The extract was washed with brine, dried (MgSO₄) and concentrated in vacuo to give 589 mg (quantitative) of 10, ν_{max} 3450 (s), 1740 (m) cm⁻¹. This was employed in the next step without further purification.

2) (2*S*,3*S*)-2-Ethyl-3-hydroxy-2-methylcyclohexanone ethylene acetal 5c. To a soln of 10 (589 mg) in diethylene glycol (50 ml) was added N₂H₄·H₂O (460 mg, 9.2 mmol) and KOH (462 mg, 85%, 7.01 mmol), and the mixture was stirred and heated at 120°C for 30 min. Then the temp was raised to 200°C and the stirring was continued for 1 h. After the mixture was cooled to room temp, this was poured into water and extracted with ether. The ether soln was washed with sat NH₄Cl soln and brine, dried (MgSO₄) and concentrated in vacuo to give 300 mg (58.8% from 10) of 5c. Its NMR and IR spectra were identical to those of the less polar isomer obtained by the acetalization of 5a and 6a. This was employed in the next step without further purification.

3) Deprotection of ethylene acetal. p-TsOH·H₂O (5 mg) was added to a soln of 5c (45 mg, 0.23 mmol) in acetone (20 ml) at room temp and the mixture was stirred for 2 h at room temp. Then this was poured into sat NaHCO₃ soln and extracted with ether. The ether soln was washed with brine, dried (MgSO₄) and concentrated in vacuo to give 32 mg (91%) of 5a, [α]_D²⁵ +77.1° (c=0.626, CHCl₃); ν_{max} 3450 (s), 1695 (s), 1060 (s), 990 (s) cm⁻¹; δ (100 MHz, CDCl₃) 0.89 (3H, t, J=8 Hz), 1.15 (3H, s), 1.35-2.20 (7H, m), 2.37 (2H, deformed t, J=7 Hz), 3.72 (1H, dd, J=6, 9 Hz). The optical purity was determined as 98.3% e.e. in the same manner as above. The sign of optical rotation and ¹H NMR spectrum of this 5a was identical with that of 5a obtained by the reduction with P. terricola. Therefore the absolute configuration of 5a from P. terricola was unambiguously determined to be (2*S*,3*S*).

Reduction with P. terricola KI 0117 in a preparative scale. 4a (8.2 g, 53 mmol) in Triton X-100 (0.2%, 80 ml) was reduced with the wet cells of P. terricola KI 0117 (87 g) in glucose soln (total 175 g/1000 ml) at 37°C for 36 h to give 7.1 g (46 mmol, 86%) of ketol; b.p. 109-114°C/5 Torr; n_D²⁴ 1.4743; [α]_D²⁴ +65.6° (c=1.07, CHCl₃); ν_{max} 3450 (s), 1695 (s), 1060 (s), 990 (s) cm⁻¹; δ (100 MHz, CDCl₃) 0.89 (3H, t, J=8 Hz), 1.15 (3H, s), 1.35-2.20 (7H, m), 2.37 (2H, deformed t, J=7 Hz), 3.72 (1H, dd, J=6, 9 Hz); MS m/z 156 (M⁺).

(2*S*,3*S*)-2-Ethyl-2-methyl-3-(3',5'-dinitro)benzoyloxy-cyclohexanone 5d. 3,5-Dinitrobenzoyl chloride (65 g, 0.28 mol) was added to a stirred and cooled soln of 5a (40 g, 0.26 mol) in pyridine (60 ml) and CH₂Cl₂ (120 ml). After the mixture was stirred for 3 h at 0°C, the reaction was quenched by adding ice and the stirring was continued for 10 min. Then the mixture was poured into dilute HCl and extracted with CH₂Cl₂. The extract was washed with CuSO₄ soln, water, sat NaHCO₃ soln and brine, dried (MgSO₄) and concentrated in vacuo to give 120 g of crude 5d. This was recrystallized from n-hexane-EtOAc (1:1) to give 65 g (72%) of pure 5d as pale yellow needles, m.p. 150.5-151.4°C; [α]_D²⁴ +72.3° (c=1.01, CHCl₃); ν_{max} 1725 (s), 1710 (s), 1630 (w), 1550 (s), 1345 (s) cm⁻¹; δ (100 MHz, CDCl₃) 0.84 (3H, t, J=8.2 Hz), 1.20 (3H, s), 1.60-2.62 (8H, m), 5.26 (1H, dd, J=5.0, 7.0 Hz), 9.13 (2H, d, J=2.1 Hz), 9.25 (1H, dd, J=2.1, 2.2 Hz). (Found: C, 54.94; H, 5.39; N, 8.14. Calc for C₁₆H₁₈O₇N₂: C, 54.85; H, 5.18; N, 8.00%).

A small portion of it was carefully saponified (0°C, 5 min) to give almost pure 5a (>99% e.e.), [α]_D²⁴ +79.2° (c=1.00, CHCl₃). This value was in good accord with that (+77.1°) of 5a obtained from the lactone 8. Therefore 5a freshly obtained by the action of P. terricola which showed smaller [α]_D value ([α]_D +65.6°) must have been contaminated with a small amount of impurities of microbial origin.

(5*S*,6*S*)-6-Methyl-5-(3',5'-dinitro)benzoyloxy-6-octanolide 11. To a stirred suspension of MCPBA (80%, 2.6 g, 12 mmol) and NaHCO₃ (1.0 g, 12 mmol) in dry CH₂Cl₂ (100 ml) was added dropwise a soln of 5d (3.5 g, 10 mmol) in dry CH₂Cl₂ (50 ml) and the mixture was stirred for 72 h at room temp. Then the mixture was filtered through a pad of celite and filter-cake was washed with CH₂Cl₂. The combined filtrate and washings were washed with 10% NaHSO₃ soln, sat NaHCO₃ soln and brine, dried (MgSO₄) and concentrated in vacuo to give 4.0 g (quantitative) of crystalline crude 11. This was employed in the next step without further purification. A small portion was recrystallized from n-hexane-EtOAc (2:1) to give an analytical sample as pale yellow needles, m.p. 160.0-161.0°C; [α]_D²⁴ +8.35° (c=1.05, CHCl₃); ν_{max} 1720 (s), 1635 (w), 1555 (s), 1350 (s) cm⁻¹; δ (100 MHz, CDCl₃) 1.01 (3H, t, J=8.0 Hz), 1.56 (3H, s), 1.60-2.33 (6H, m), 2.70-2.92 (2H, m), 5.42 (1H, t, J=5.0 Hz), 9.12 (2H, d, J=2.1 Hz), 9.25 (1H, dd, J=2.1, 2.2 Hz). (Found: C, 52.64; H, 4.88; N, 7.74. Calc for C₁₆H₁₈O₈N₂: C, 52.45; H, 4.95; N, 7.65%).

Methyl (5*S*,6*S*)-5,6-isopropylidenedioxy-6-methyloctanoate 12. K₂CO₃ (40 mg, 0.29 mmol) was added to a stirred soln of crude 11 (4.0 g, 10 mmol) in MeOH (10 ml) and CH₂Cl₂ (30 ml) and the mixture was stirred for 30 min at room temp. To this was added p-TsOH until the color of the soln changed from red to yellow. Then most of the solvent was removed by evaporation, the residue was diluted with acetone and filtered through a pad of celite. The filter-cake was washed with acetone and the combined filtrate and washings were concentrated in vacuo until the volume was ca. 20 ml. To this was added 2,2-dimethoxypropane (5 ml, 41 mmol) and the mixture was stirred for 3 h at room temp. Then a half of the solvent was removed by evaporation and the residual soln was poured into sat NaHCO₃ soln and extracted with ether. The ether soln was washed with brine, dried (MgSO₄) and concentrated in vacuo to give a mixture of 12 and crystalline methyl 3,5-dinitrobenzoate. The following operation was repeated three times to remove crystalline methyl 3,5-dinitrobenzoate; dilution of the mixture with n-hexane, filtration, concentration of the filtrate. The oily residue was distilled to give 2.2 g (90% from 5d) of 12, b.p. 83°C/0.55 Torr; n_D²⁴ 1.4333; [α]_D²⁴ -13.9° (c=1.07, MeOH); ν_{max} 1740 (s), 1260 (s), 1220 (s), 1180 (br.s) cm⁻¹; δ (100 MHz, CDCl₃) 0.95 (3H, t, J=7.6 Hz), 1.06 (3H, s), 1.33 (3H, s), 1.43 (3H, s), 1.25-2.05 (6H, m), 2.38 (2H, deformed t, J=7.0), 3.68 (3H, s), 3.70-3.82 (1H, m); GLC (PEG 0.2 mm x 50 m, 1.2 kg/cm², 170°C) Rt 13.6 min. (Found: C, 63.68; H, 9.78. Calc for C₁₃H₂₄O₄: C, 63.90; H, 9.90%).

A mixture of (±)-12 and its diastereomer (±)-13 was prepared from a mixture of 5a, 5a', 6a and 6a'. 100 MHz ¹H NMR spectra of this mixture showed signals at 1.06 ppm and 1.18 ppm due to C-6 Me singlet.²² The ester 12 obtained above showed only 1.06 ppm singlet, therefore 12 was revealed to be diastereomerically pure.

(5*S*,6*S*)-5,6-Isopropylidenedioxy-6-methyl-1-octanol 14. To a stirred and cooled suspension of LAH (0.76 g, 20 mmol) in dry ether (10 ml) was added dropwise a soln of 12 (3.9 g, 16 mmol) in dry ether (20 ml) at 0°C and the mixture was stirred for 2.5 h at room temp. It was again cooled to 0°C and the reaction was quenched by the addition of water (0.76 ml), 15% NaOH soln (0.76 ml) and water (2.28 ml) dropwise. Anhydrous Na₂SO₄ was added to the mixture and it was stirred for 12 h at room temp, then filtered through a pad of celite and the filter-cake was washed with THF. The combined filtrate and washings were concentrated in vacuo and the residual oil was distilled to give 3.1 g (90%) of 14, b.p. 92-94°C/0.45 Torr; n_D²⁴

1,4441; $[\alpha]_D^{24}$ -7.55° (c=1.02, CHCl₃); ν_{\max} 3450 (br.s), 1250 (s), 1210 (s), 1190 (s), 1110 (s), 1060 (s), 1020 (br.s) cm⁻¹, δ (60 MHz, CCl₄) 0.90 (3H, t, J=7.2 Hz), 0.98 (3H, s), 1.22 (3H, s), 1.31 (3H, s), 1.20-2.30 (9H, s), 3.30-3.72 (3H, m). (Found: C, 66.35; H, 11.17. Calc for C₁₂H₂₄O₃: C, 66.63; H, 11.18%).

A small portion of this was treated with aq AcOH (75 v/v%) at 50°C for 2 h to remove acetonide protective group to give a triol, which was converted to the corresponding bis (R)- and (S)-MTPA esters 15. HPLC analysis; eluent: n-hexane-THF (5:1), 1.0 ml/min; Rt 11.3 min [(S)-MTPA ester], 14.2 min [(R)-MTPA ester]; bis (R)-MTPA ester of 15, Rt 14.2 min (100%). Therefore 14 was revealed to be enantiomerically pure.

(3S,4S)-3,4-Isopropylidenedioxy-3-methyl-7-octene 17. To a stirred and cooled soln of 14 (2.5 g, 12 mmol) and o-nitrophenyl selenocyanate (3.2 g, 14 mmol) in dry THF (30 ml) was added dropwise (n-Bu)₃P (3.5 ml, 2.8 g, 14 mmol) under Ar at 0°C. The mixture was stirred for 5 h at room temp and the solvent was removed by evaporation. The residue was chromatographed over SiO₂ (80 g). Elution with C₆H₆-ether (10:1) gave 5.4 g of crude 16. This was dissolved in dry THF (25 ml) and to this was added dropwise 35% H₂O₂ (8.7 ml) at 0-5°C with stirring. After stirring for 2.5 h at room temp, the mixture was poured into sat NaHCO₃ soln and extracted with ether. The ether soln was washed with 5% Na₂S₂O₃ soln, 10% Na₂CO₃ soln and brine, dried (MgSO₄) and concentrated *in vacuo*. The residual oil was filtered through a column of neutral Al₂O₃ (grade IV). The fraction eluted with n-pentane was concentrated *in vacuo* followed by distillation to give 1.8 g (79%) of 17, b.p. 130-150°C(bath)/15 Torr; n_D^{24} 1.4286; $[\alpha]_D^{26}$ -1.25° (c=1.02, CHCl₃); ν_{\max} 1640 (w), 1250 (s), 1210 (s), 1190 (s), 920 (s), 870 (s) cm⁻¹; δ (60 MHz, CCl₄) 0.90 (3H, t, J=6.3 Hz), 0.96 (3H, s), 1.20 (3H, s), 1.30 (3H, s), 1.15-2.40 (6H, m), 3.55 (1H, dd, J=4.1, 8.7 Hz), 4.70-5.12 (2H, m), 5.75 (1H, ddt, J=10.3, 18.0, 6.0 Hz). (Found: C, 72.49; H, 10.86. Calc for C₁₂H₂₂O₂: C, 72.68; H, 11.18%).

(3S,4S)-3,4-Isopropylidenedioxy-3-methyl-7-octyne 19. Br₂ (1.3 g, 8.1 mmol) was added to a stirred and cooled soln of 17 (1.6 g, 8.1 mmol) in CCl₄ (20 ml) and the mixture was stirred for 2 h at 0-10°C. Then it was poured into 5% Na₂S₂O₃ soln and extracted with ether. The ether soln was washed with 5% NaHCO₃ soln and brine, dried (CaCl₂) and concentrated *in vacuo* to give 2.8 g of dibromide 18. This was dissolved in dry THF (20 ml) and this was added to a suspension of NaNH₂ (from 2.3 g of Na; 11 eq) in liq NH₃ (100 ml) dropwise at -60°C. After stirring for 3 h at -60-70°C, the temp was gradually raised to ca. -33°C with stirring, then the reaction was quenched by adding solid NH₄Cl (5.0 g, 93 mmol) followed by sat NH₄Cl soln and stirred overnight. The mixture was extracted with ether and the ether soln was washed with water and brine, dried (MgSO₄) and concentrated *in vacuo*. The residual oil was distilled to give 0.80 g (50%) of 19, b.p. 86-91°C/15 Torr; n_D^{24} 1.4396; $[\alpha]_D^{24.5}$ -33.5° (c=1.03, CHCl₃); ν_{\max} 3320 (m), 2120 (w), 1255 (s), 1215 (s), 1000 (s) cm⁻¹; δ (60 MHz, CCl₄) 0.92 (3H, t, J=6.4 Hz), 0.98 (3H, s), 1.22 (3H, s), 1.30 (3H, s), 1.10-2.50 (7H, m), 3.70 (1H, dd, J=4.0, 8.7 Hz). (Found: C, 73.07; H, 10.01. Calc for C₁₂H₂₀O₂: C, 73.43; H, 10.27%).

Methyl (6S,7S)-6,7-isopropylidenedioxy-7-methyl-2-nonynoate 20. To a stirred and cooled soln of 19 (3.8 g, 19 mmol) in dry THF (60 ml) was added a soln of n-BuLi (1.6 N in n-hexane, 16 ml, 26 mmol) dropwise at 5°C and the mixture was stirred for 1 h at 0°C under Ar. Then the soln was cooled to -78°C and a soln of ClCO₂Me (2.5 g, 26 mmol) in dry THF (6 ml) was added dropwise and the mixture was stirred for 1 h at -78°C. It was then stirred at room temp for additional 2.5 h. The reaction was quenched by adding sat NH₄Cl soln, and the mixture was extracted with ether. The ether soln was washed with sat NaHCO₃ soln and brine, dried (MgSO₄) and concentrated *in vacuo*. The residual oil was chromatographed over SiO₂ (70 g). Elution with n-hexane-ether (20:1) gave recovered 19 (0.45 g). Further elution with n-hexane-ether (20:1-10:1) gave 3.7 g (86% based on the consumed 19) of 20. A small portion of it was distilled to give an analytical sample, b.p. 115-118°C/5 Torr; n_D^{25} 1.4536; $[\alpha]_D^{26}$ -38.7° (c=0.995, CHCl₃); ν_{\max} 2250 (m), 1720 (s), 1255 (s) cm⁻¹; δ (60 MHz, CCl₄) 0.92 (3H, t, J=6.4 Hz), 0.99 (3H, s), 1.24 (3H, s), 1.30 (3H, s), 1.12-2.70 (6H, m), 3.65, (3H, s), 3.55-3.85 (1H, m). (Found: C, 65.62; H, 8.42. Calc for C₁₄H₂₂O₄: C, 66.11; H, 8.72%).

Methyl (2Z,6S,7S)-6,7-isopropylidenedioxy-7-methyl-3-phenylthio-2-nonenolate 21 and its (2E,6S,7S)-isomer 22. NaOH (0.69 g, 17 mmol) was added to a stirred soln of C₆H₅SH (1.9 g, 17 mmol) in MeOH (30 ml) and the mixture was stirred for 30 min at room temp. To this was added dropwise a soln of 20 (3.7 g, 15 mmol) in MeOH (10 ml) and the stirring was continued for 4 h at room temp. A soln of AcOH (1 g, mmol) in water (5 ml) was added to quench the reaction. The reaction mixture was diluted with ether and it was washed with 4% NaOH soln and brine, dried (MgSO₄) and concentrated *in vacuo*. The residual oil (5.0 g) was repeatedly chromatographed over SiO₂. Elution with n-hexane-ether (40:1) gave 0.58 g (11%) of 22, n_D^{23} 1.5253; $[\alpha]_D^{23}$ +18.3° (c=1.10, MeOH); ν_{\max} 3100 (w), 1710 (s), 1600 (s), 1370 (m), 1340 (m), 1170 (br.s), 750 (m), 710 (s), 690 (m), 680 (m) cm⁻¹; δ (100 MHz, CDCl₃) 0.98 (3H, t, J=7.8 Hz), 1.12 (3H, s), 1.34 (3H, s), 1.46 (3H, s), 1.14-2.20 (4H, m), 3.63 (3H, s), 3.88 (1H, dd, J=4.5, 8.4 Hz), 5.17 (1H, s), 7.30-7.53 (5H, m). (Found: C, 65.86; H, 7.83. Calc for C₂₀H₂₈O₄S: C, 65.90; H, 7.74 %). Further elution gave 4.0 g (76%) of 21, $n_D^{24.5}$ 1.5264; $[\alpha]_D^{24.5}$ +9.25° (c=1.02, MeOH); ν_{\max} 3100 (w), 1720 (s), 1590 (s), 1380 (s), 1200 (br.s), 760 (m), 710 (m), 700 (m) cm⁻¹; δ (100 MHz, CDCl₃) 0.85 (3H, t, J=7.8 Hz), 0.92 (3H, s), 1.20 (3H, s), 1.33 (3H, s), 1.00-1.82 (4H, m), 2.00-2.64 (2H, m), 3.30 (1H, dd, J=4.5, 8.4 Hz), 3.65 (-OMe singlet of the trace of the desconjugated ester), 3.78 (3H, s), 5.94 (1H, s), 7.20-7.65 (5H, m). (Found: C, 65.70; H, 7.59. Calc for C₂₀H₂₈O₄S: C, 65.90; H, 7.74%).

Methyl (2E,6S,7S)-3-ethyl-6,7-isopropylidenedioxy-7-methyl-2-nonenolate 23. To a stirred and cooled suspension of CuI (2.1 g, 11 mmol) in dry THF (3 ml) was added a soln of EtMgBr (27 mmol in 20 ml THF) at -60-70°C and the stirring was continued for 10 min under Ar. To the mixture was added a soln of 21 (2.0 g, 5.5 mmol) in dry THF (30 ml). After stirring for 2 h at -78°C, the reaction was quenched by adding sat NH₄Cl soln. The stirring was continued for additional 1 h at room temp. Then the mixture was extracted with ether. The ether soln was washed with sat NH₄Cl soln and 5% NaOH soln. The precipitated inorganic material was removed by filtration. The ether layer was washed with brine, dried (MgSO₄) and concentrated *in vacuo*. The residual oil was chromatographed over SiO₂ (40 g). Elution with n-hexane-ether (20:1) gave 1.3 g (83%) of 23. A small portion of it was distilled to give an analytical sample, b.p. 68-70°C/0.25 Torr; n_D^{22} 1.4576; $[\alpha]_D^{22}$ -11.7° (c=0.890, MeOH); ν_{\max} 1720 (s), 1645 (m), 1380 (m), 1210 (s), 1150 (s) cm⁻¹; δ (100 MHz, CDCl₃) 0.97 (3H, t, J=7.1 Hz), 1.08 (3H, s), 1.11 (3H, t, J=7.1 Hz), 1.33 (3H, s), 1.43 (3H, s), 1.40-1.90 (4H, m), 2.00-2.84 (4H, m), 3.60-3.85 (1H, m), 3.70 (3H, s), 5.66 (1H, br.s); GLC (5% PEG-20M; 5 mm x 1.5 m; 150°C; 1.0 kg/cm²): Rt 6.1 min (99.7%). (Found: C, 67.51; H, 9.91. Calc for C₁₆H₂₆O₄: C, 67.57; H, 9.93%).

Methyl (2E,6S,7S)-6,7-isopropylidenedioxy-3,7-dimethyl-2-nonenolate 24. In the same manner as above, 21 (2.0 g, 5.5 mmol) was converted to 24 (1.2 g, 81%), 89-92°C/1 Torr; $n_D^{24.5}$ 1.4562; $[\alpha]_D^{24.5}$ -21.3° (c=0.960, MeOH); ν_{\max} 1720 (s), 1650 (m), 1375 (m), 1220 (s), 1155 (s) cm⁻¹; δ (100 MHz, CDCl₃) 0.96 (3H, t, J=7.8 Hz), 1.08 (3H, s), 1.32 (3H, s), 1.43 (3H, s),

1.45-1.90 (4H, m), 2.19 (3H, d, J=1.2 Hz), 1.99-2.60 (2H, m), 3.70 (3H, s), 3.60-3.80 (1H, m), 5.73 (1H, q, J=1.2 Hz); GLC (10% PEG-20M; 5 mm x 2 m; 150°C + 3°C/min; N₂, 1.1 kg/cm²): Rt 14.2 min (100%). (Found: C, 66.24; H, 9.59. Calc for C₁₅H₂₆O₄: C, 66.63; H, 9.69%).

(2E,6S,7S)-3-Ethyl-6,7-isopropylidenedioxy-7-methyl-2-nonen-1-ol 25. To a stirred and cooled soln of 23 (562 mg, 1.96 mmol) in dry toluene (5 ml) was added dropwise a soln of DIBAL-H (1 M in n-hexane, 4.2 ml, 4.2 mmol) at 0°C under Ar. After stirring for 30 min at 0°C, the reaction was quenched by adding MeOH (2 ml). The stirring was continued for 10 min at room temp. Then the mixture was filtered through a pad of celite and the filter-cake was washed with MeOH. The combined filtrate and washings were concentrated in vacuo to give 523 mg (quantitative) of 25, n_D²⁴ 1.4610; [α]_D²⁴ -3.20° (c=0.950, CHCl₃); ν_{max} 3400 (br.m), 1660 (w), 1210 (s), 1000 (s) cm⁻¹; δ (100 MHz, CDCl₃) 0.96 (3H, t, J=7.8 Hz), 1.00 (3H, t, J=7.8 Hz), 1.08 (3H, s), 1.33 (3H, s), 1.43 (3H, s), 1.45-2.50 (9H, m), 3.72 (1H, dd, J=3.7, 8.3 Hz), 4.19 (2H, br.d, J=6.9 Hz), 5.40 (1H, br.t, J=6.9 Hz). (Found: C, 69.78; H, 10.89. Calc for C₁₅H₂₈O₃: C, 70.27; H, 11.01%).

(2E,6S,7S)-6,7-Isopropylidenedioxy-3,7-dimethyl-2-nonen-1-ol 26. In almost the same manner as above, 24 (1.15 g, 4.26 mmol) was converted to 26 (930 mg, 90.2%), n_D²⁵ 1.4584; [α]_D²⁵ -5.42° (c=0.985, CHCl₃); ν_{max} 3400 (br.m), 1660 (s), 1250 (m), 1210 (s), 1000 (br.s) cm⁻¹; δ (60 MHz, CCl₄) 0.90 (3H, t, J=6.4 Hz), 0.98 (3H, s), 1.20 (3H, s), 1.30 (3H, s), 1.65 (3H, s), 1.10-2.40 (7H, m), 3.55 (1H, dd, J=4.8, 8.2 Hz), 4.00 (2H, br.d, J=7.1 Hz), 5.35 (1H, br.t, J=7.1 Hz). (Found: C, 69.28; H, 10.91. Calc for C₁₄H₂₆O₃: C, 69.38; H, 10.81%).

(3S,4S,7E)-9-Bromo-7-ethyl-3,4-isopropylidenedioxy-3-methyl-7-octene 27. n-BuLi (1.53 N in n-hexane, 0.61 ml, 0.93 mmol) was added dropwise to a stirred and cooled soln of 25 (238 mg, 0.93 mmol) and Ph₃CH (3 mg) in dry ether (1 ml) and dry HMPA (1 ml) at 0°C under Ar until the characteristic red color persisted. After stirring for 5 min at 0°C, a soln of p-TsCl (190 mg, 1.0 mmol) in dry ether (1.2 ml) was added dropwise and the mixture was stirred for 45 min at 0°C. Then anhydrous LiBr (450 mg, 5.1 mmol) was added to the mixture and it was stirred overnight at room temp. The mixture was poured into sat NaHCO₃ soln and extracted with n-hexane. The n-hexane soln was washed with water, sat NaHCO₃ soln and brine, dried (MgSO₄) and concentrated in vacuo to give 315 mg (quantitative) of 27. This was employed in the next step without further purification. ν_{max} 1655 (w), 1250 (m), 1200 (br.s), 1000 (m) cm⁻¹.

(3S,4S,7E)-9-Bromo-3,4-isopropylidenedioxy-3,7-dimethyl-7-octene 28. In the same manner as above, 26 (485 mg, 2.00 mmol) was converted to 28 (722 mg, quantitative). ν_{max} 1660 (w), 1260 (m), 1210 (s), 1000 (m) cm⁻¹.

Methyl (6E,10S,11S)-7-ethyl-10,11-isopropylidenedioxy-11-methyl-3-oxo-6-tridecanoate 29. To a stirred and cooled suspension of NaH (66 mg, 1.7 mmol, 60% dispersion in mineral oil) in dry THF (1 ml) was added dropwise a soln of methyl acetoacetate (174 mg, 1.5 mmol) in dry THF (1 ml) at 0°C under Ar. The mixture was stirred for 15 min at 0°C and n-BuLi (1.53 N in n-hexane, 0.98 ml, 1.5 mmol) in dry THF (1 ml) was added dropwise to it below 5°C. The mixture was stirred for 20 min at 0°C and a soln of 27 (300 mg, 0.94 mmol) in dry THF (2 ml) was added dropwise to it at 0°C. After stirring for 2 h at -5°C, the reaction was quenched by adding sat NH₄Cl soln, and the mixture was extracted with ether. The ether soln was washed with water and brine, dried (MgSO₄) and concentrated in vacuo. The residual oil was chromatographed over SiO₂ (6 g). Elution with n-hexane-ether (10:1-5:1) gave 245 mg (78% from 25) of 29, n_D²³ 1.4633; [α]_D²³ -4.68° (c=1.08, MeOH); ν_{max} 1755 (s), 1725 (s), 1255 (s), 1215 (s) cm⁻¹; δ (100 MHz, CDCl₃) 0.96 (3H, t, J=8.0 Hz), 0.98 (3H, t, J=8.0 Hz), 1.08 (3H, s), 1.33 (3H, s), 1.44 (3H, s), 1.30-2.68 (12H, m), 3.60-3.80 (1H, m), 3.46 (2H, s), 3.78 (3H, s), 5.07 (1H, t, J=7.5 Hz). (Found: C, 67.85; H, 9.63. Calc for C₂₀H₃₄O₅: C, 67.76; H, 9.67%).

Methyl (2E,6E,10S,11S)-7-ethyl-10,11-isopropylidenedioxy-3,11-dimethyl-2,6-tridecadienoate 33. To a stirred and cooled suspension of NaH (20 mg, 0.50 mmol, 60% dispersion in mineral oil) in dry THF (1 ml) was added dropwise a soln of 29 (140 mg, 0.395 mmol) in dry THF (1.5 ml) at 0°C under Ar. The mixture was stirred at 0°C for 15 min and (EtO)₂P(O)Cl (72 μl, 86 mg, 0.50 mmol) was added dropwise at 0°C. After stirring for 2 h at room temp, the mixture was poured into sat NH₄Cl soln and extracted with ether. The ether soln was washed with sat NaHCO₃ soln and brine, dried (MgSO₄) and concentrated in vacuo to give 233 mg of crude phosphoric ester 31. This was dissolved in dry ether (1.5 ml) and the ether soln was added dropwise to a soln of Me₂QuLi (1.40 mmol) which was prepared according to the reported procedure²⁶ at -60°C under Ar. The mixture was stirred for 1.5 h at -70°C, and quenched by adding sat NH₄Cl soln. The stirring was continued for additional 5 min. Then the mixture was extracted with ether. The ether soln was washed with sat NaHCO₃ soln and brine, dried (MgSO₄) and concentrated in vacuo. The residual oil was chromatographed over SiO₂ (3 g). Elution with n-hexane-ether (20:1) gave 83.0 mg of crude 33. The purity of it was 78.1% by GLC analysis. (5% PEG-20M; 5 mm x 1.5 m; 200°C; N₂; Rt 4.9 min). Further purification was carried out by using Merck Lobar® column. Pure 33 (118 mg, 35%) was obtained from 199 mg of crude 33. GLC (same condition): Rt 4.9 min (94.7 %); HPLC [eluent: n-hexane-THF (30:1)]: Rt 12.9 min (100%). n_D²³ 1.4677; [α]_D²³ -4.15° (c=1.11, MeOH); ν_{max} 1720 (s), 1645 (m), 1370 (m), 1250 (m), 1220 (s), 1140 (s), 1000 (m) cm⁻¹; δ (100 MHz, CDCl₃) 0.96 (3H, t, J=7.8 Hz), 0.98 (3H, t, J=8.3 Hz), 1.07 (3H, s), 1.33 (3H, s), 1.44 (3H, s), 1.35-1.70 (4H, m), 2.18 (3H, d, J=1.3 Hz), 1.80-2.32 (8H, m), 3.65-3.80 (1H, m), 3.70 (3H, s), 5.00-5.18 (1H, m), 5.68 (1H, br.s). (Found: C, 71.84; H, 10.17. Calc for C₂₁H₃₆O₄: C, 71.55; H, 10.30%).

Methyl (2E,6E,10S,11S)-10,11-isopropylidenedioxy-3,7,11-trimethyl-2,6-tridecadienoate 34. In almost the same manner as above, 26 (485 mg, 2.00 mmol) was converted to 34 (110 mg, 16.3%). n_D²⁴ 1.4718; [α]_D²⁴ -7.33° (c=1.02, MeOH); ν_{max} 1720 (s), 1645 (m), 1370 (m), 1250 (m), 1220 (s), 1145 (s) cm⁻¹; δ (100 MHz, CDCl₃) 0.96 (3H, t, J=7.8 Hz), 1.07 (3H, s), 1.32 (3H, s), 1.44 (3H, s), 1.63 (3H, d, J=1.2 Hz), 1.26-1.70 (4H, m), 1.80-2.30 (6H, m), 3.70 (3H, s), 3.62-3.82 (1H, m), 5.05-5.25 (1H, m), 5.68 (1H, br.s); GLC (5% PEG-20M, 5 mm x 1.5 m; 170°C; N₂, 1.0 kg/cm²; Rt 12.4 min (1.7%), 16.5 min (98.3%). (Found: C, 70.53; H, 9.86. Calc for C₂₀H₃₄O₄: C, 70.97; H, 10.13%).

Methyl (2E,6E,10S,11S)-7-ethyl-10,11-dihydroxy-3,11-dimethyl-2,6-tridecanoate 35a. A soln of 33 (112 mg, 0.318 mmol) in 75% (v/v) aq AcOH (4 ml) was stirred and heated for 6 h at 50°C. After the mixture was cooled to 0°C, this was diluted with EtOAc and a soln of NaOH (2.00 g, 50.0 mmol) in water (10 ml) was added dropwise to this. This was poured into brine and extracted with EtOAc. The EtOAc soln was washed with sat NaHCO₃ soln and brine, dried (Na₂SO₄) and concentrated in vacuo to give 98.0 mg (98.8%) of 18a. This was employed in the next step without further purification. [α]_D²⁵ -17° (c=0.99, MeOH); ν_{max} 3450 (br.m), 1720 (s), 1645 (m), 1220 (s), 1150 (s) cm⁻¹. This was revealed to be enantiomerically pure by the HPLC analysis of the corresponding MTPA esters: eluent: n-hexane-THF (5:1); 35b and 35c, Rt 6.84 and 7.61 min; 35b, 7.61 min (100%).

Methyl (2E,6E,10E,11E)-10,11-dihydroxy-3,7,11-trimethyl-2,6-tridecadienoate 36a. In the same manner as above, 34 (106 mg, 0.314 mmol) was converted to 36b (100 mg, quantitative), $[\alpha]_D^{24} -20^\circ$ ($c=0.96$, MeOH); ν_{\max} 3455 (br,m), 1720 (s), 1650 (m), 1220 (s), 1150 (s) cm^{-1} . This was revealed to be enantiomerically pure by the HPLC analysis of the corresponding NTPA esters: eluent: *n*-hexane-THF (7:1); 36b and 36c, Rt 10.4 and 12.5 min; 36b, 12.5 min (100%).

Methyl (2E,6E,10E,11E)-10,11-epoxy-7-ethyl-3,11-dimethyl-2,6-tridecanoate (JH I) 1. To a stirred and cooled soln of 35a (50 mg, 0.16 mmol) in dry CH_2Cl_2 (1.5 ml) and Et_3N (24.0 mg, 0.238 mmol) was added solid Mg_2O (36 mg, 0.21 mmol) at 0°C . After stirring for 1 h at 0°C , the mixture was poured into sat NaHCO_3 soln and extracted with EtOAc . The EtOAc soln was washed with brine, dried (Na_2SO_4) and concentrated *in vacuo* to give 55.7 mg of 37. This was dissolved in MeOH (1 ml) and to this was added NaOMe (28.9 mg, 0.150 mmol, 28% in MeOH) in MeOH (1 ml) dropwise at 0°C with stirring. After the mixture was stirred for 10 min at 0°C , it was poured into sat NaHCO_3 soln, and extracted with ether. The ether soln was washed with NaHCO_3 soln and brine, dried (MgSO_4) and concentrated *in vacuo*. The residual oil was purified by prep TLC (Merck Kiesel gel 60 F-254) to give 37 mg (79%) of pure 1, $n_D^{24} 1.4732$; $[\alpha]_D^{24} +14.9^\circ$ ($c=0.935$, CHCl_3); $[\alpha]_D^{25} +14.5^\circ$ ($c=0.78$, MeOH); ν_{\max} 2990 (s), 2950 (m), 2895 (m), 1720 (s), 1650 (m), 1470 (m), 1450 (m), 1435 (m), 1380 (m), 1360 (m), 1325 (w), 1280 (w), 1250 (w), 1225 (s), 1150 (s), 1060 (w), 1030 (w), 920 (w), 880 (m), 860 (m), 820 (m), 785 (w), 730 (w) cm^{-1} ; δ (400 MHz, CCl_4) 0.98 (3H, t, $J=7.8$ Hz), 0.98 (3H, t, $J=7.8$ Hz), 1.19 (3H, s), 1.35-1.63 (4H, m), 2.13 (3H, d, $J=1.2$ Hz), 1.99-2.23 (8H, m), 2.50 (1H, dd, $J=5.1, 7.3$ Hz), 3.61 (3H, s), 5.05 (1H, br,t, $J=6.4$ Hz), 5.57 (1H, br,d, $J=1.2$ Hz); ^{13}C NMR δ 9.65, 13.10, 18.84, 21.59, 23.17, 25.57, 25.80, 27.26, 33.29, 41.19, 50.72, 61.78, 64.65, 115.31, 122.91, 141.28, 159.83, 167.14. HPLC: eluent, *n*-hexane-THF (30:1); Rt 12.9 min (100%). (Found: C, 73.25; H, 10.08. Calc for $\text{C}_{18}\text{H}_{30}\text{O}_3$: C, 73.43; H, 10.27 %).

Methyl (2E,6E,10E,11E)-10,11-epoxy-3,7,11-trimethyl-2,6-tridecadienoate (JH II) 2. In the same manner as above, 36a (80 mg, 0.27 mmol) was converted to 2 (50 mg, 66%), $n_D^{25} 1.4774$; $[\alpha]_D^{25} +17.6^\circ$ ($c=0.590$, MeOH); ν_{\max} 3000 (s), 2950 (s), 2900 (m), 2850 (m), 1725 (s), 1650 (s), 1470 (m), 1450 (m), 1440 (m), 1420 (w), 1385 (m), 1360 (m), 1325 (w), 1280 (w), 1225 (s), 1150 (s), 1110 (w), 1060 (m), 1030 (m), 1000 (w), 980 (w), 920 (w), 880 (m), 865 (m), 800 (w), 740 (w) cm^{-1} ; δ (400 MHz, CCl_4) 0.98 (3H, t, $J=7.8$ Hz), 1.19 (3H, s), 1.62 (3H, s), 1.34-1.65 (4H, m), 2.14 (3H, d, $J=1.3$ Hz), 2.01-2.21 (6H, m), 2.49 (1H, dd, $J=5.1, 7.4$ Hz), 3.61 (3H, s), 5.11 (1H, br,t, $J=5.9$ Hz), 5.57 (1H, br,s); ^{13}C NMR δ 9.65, 16.03, 18.78, 21.59, 25.80, 25.92, 27.03, 36.51, 40.78, 50.72, 61.72, 64.53, 115.31, 123.44, 135.38, 159.89, 167.20. HPLC: eluent: *n*-hexane-THF (60:1), Rt 21.6 min (100%). (Found: C, 72.93; H, 9.89. Calc for $\text{C}_{17}\text{H}_{28}\text{O}_3$: C, 72.82; H, 10.06%).

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REFERENCES

- H. Röllner, K.-H. Dahm, C. C. Sweeley and B. M. Trost, *Angew. Chem. Int. Ed. Engl.* **6**, 179 (1967).
- a) A. S. Meyer, H. A. Schneiderman, E. Hanzmann and J. H. Ko, *Proc. Natl. Acad. Sci. USA* **60**, 853 (1968). b) A. S. Meyer, E. Hanzmann, H. A. Schneiderman, L. I. Gilbert and M. Boyette, *Arch. Biochem. Biophys.* **137**, 190 (1970).
- K. J. Judy, D. A. Schooley, L. L. Dunham, M. S. Hall, B. J. Bergot and J. B. Siddall, *Proc. Natl. Acad. Sci. USA* **70**, 1509 (1973).
- B. J. Bergot, G. C. Jamieson, M. A. Ratcliff and D. A. Schooley, *Science* **210**, 336 (1980).
- T. Koyama, K. Ogura, F. C. Baker, G. C. Jamieson and D. A. Schooley, *J. Am. Chem. Soc.* **109**, 2853 (1987).
- Reviews: a) D. A. Schooley, "Analysis of the Naturally Occurring Juvenile Hormones -- Their Isolation, Identification, and Titer Determination at Physiological Levels" in *Analytical Biochemistry of Insects* (R. B. Turner, Ed.), pp. 241-287, Elsevier, Amsterdam 1977. b) K. Mori, "Synthetic Chemistry of Insect Pheromones and Juvenile Hormones" in *Recent Developments in the Chemistry of Natural Carbon Compounds* (R. Bognár, V. Bruckner and Cs. Szántay, Ed.), vol. 9, pp. 9-209, Akadémiai Kiadó, Budapest 1979. c) *Regulation of Insect Development and Behavior*, Part 1, pp. 147-244, Wrocław Technical University Press, Wrocław 1981.
- A. S. Meyer and E. Hanzmann, *Biochem. Biophys. Res. Commun.* **41**, 891 (1970).
- A. S. Meyer, E. Hanzmann and R. C. Murphy, *Proc. Natl. Acad. Sci. USA* **68**, 2312 (1971).
- P. Loew and W. S. Johnson, *J. Am. Chem. Soc.* **93**, 3765 (1971).
- D. J. Faulkner and M. R. Petersen, *J. Am. Chem. Soc.* **93**, 3766 (1971).
- K. Nakanishi, D. A. Schooley, M. Koreeda and J. Dillon, *Chem. Commun.* 1235 (1971).
- G. D. Prestwich and C. Wawrzynczyk, *Proc. Natl. Acad. Sci. USA* **82**, 5290 (1985).
- K. Inai, S. Marumo and K. Mori, *J. Am. Chem. Soc.* **96**, 5925 (1974).
- K. Mori and H. Mori, *Tetrahedron*, in the press.
- M. E. Garst and B. J. McBride, *J. Org. Chem.* **48**, 1362 (1983).
- K. Mori and H. Mori, *Tetrahedron* **42**, 5531 (1986) and refs. cited therein.
- D. W. Brooks, H. Mazdiyani and P. G. Grothaus, *J. Org. Chem.* **52**, 3223 (1987).
- T. Sugai, unpublished results.
- J. A. Dale and H. S. Mosher, *J. Am. Chem. Soc.* **95**, 512 (1973).
- T. Sugai, M. Fujita and K. Mori, *Nippon Kagaku Kaishi* 1315 (1983).
- K. Mori, H. Mori and T. Sugai, *Tetrahedron* **41**, 919 (1985).
- C. W. K. Cavill, D. G. Laing and P. J. Williams, *Aust. J. Chem.* **22** 2145 (1969).
- P. A. Grieco, S. Gilman and M. Nishizawa, *J. Org. Chem.* **41**, 1845 (1976).
- a) S. Kobayashi and T. Mukaiyama, *Chem. Lett.* 705 (1974). b) S. Kobayashi and T. Mukaiyama, *Chem. Lett.* 1425 (1974). c) T. Mukaiyama, H. Toda and S. Kobayashi, *Chem. Lett.* 535 (1975).
- G. Stork, P. A. Grieco and M. Gregson, *Tetrahedron Lett.* 1393 (1969).
- P. W. Sum and L. Weiler, *Can. J. Chem.* **57**, 1431 (1979).
- For the ^1H NMR spectrum of (\pm)-JH II, see: K. Mori, A. Sato and M. Matsui, *Agric. Biol. Chem.* **36**, 1931 (1972).
- For the ^1H NMR spectrum of the C-10 epimer (10-*trans*-epoxide) of (\pm)-JH II, see: K. Mori, *Agric. Biol. Chem.* **36**, 2563 (1972).
- B. M. Trost, *Acc. Chem. Res.* **3**, 122 (1970).